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NEWS 18 Aug 05 NTIS has been reloaded and enhanced
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NEWS 23 Sep 04 CAPIO has been reloaded and enhanced
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L1 5913969 GENETIC(W) MATERIAL OR DNA OR RNA (2A) EXTRACT? OR ISOLAT?

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L2 305809 L1 (S) COLUMN OR IMMOBILIZ?

=> s l2 (s) label?
L3 18648 L2 (S) LABEL?

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L5 4 L3 (S) RADICAL(W) MEDIAT?

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L5 ANSWER 1 OF 4 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.
AN 1999:119009 BIOSIS
DN PREV199900119009
TI Increased RNA oxidation and decreased levels of repair products in Alzheimer's disease ventricular CSF.
AU Lovell, Mark A.; Gabbita, S. Prasad; Markesbery, William R. (1)
CS (1) 101 Sanders-Brown Bldg., Univ. Kentucky, Lexington, KY 40536-0230, USA
SO Journal of Neurochemistry, (Feb., 1999) Vol. 72, No. 2, pp. 771-776.
ISSN: 0022-3042.
DT Article

LA English

L5 ANSWER 2 OF 4 MEDLINE
AN 1999127942 MEDLINE
DN 99127942 PubMed ID: 9930752
TI Increased DNA oxidation and decreased levels of repair products in Alzheimer's disease ventricular CSF.
AU Lovell M A; Gabbita S P; Markesbery W R
CS Sanders-Brown Center on Aging, and Department of Chemistry, University of Kentucky, Lexington 40536-0230, USA.
NC 1P01-AG05119 (NIA)
5P01-AG05144 (NIA)
SO JOURNAL OF NEUROCHEMISTRY, (1999 Feb) 72 (2) 771-6.
Journal code: 2985190R. ISSN: 0022-3042.
CY United States
DT Journal; Article; (JOURNAL ARTICLE)
LA English
FS Priority Journals
EM 199903
ED Entered STN: 19990129
Last Updated on STN: 19990223
Entered Medline: 19990211

L5 ANSWER 3 OF 4 EMBASE COPYRIGHT 2002 ELSEVIER SCI. B.V.
AN 1999042164 EMBASE
TI Increased DNA oxidation and decreased levels of repair products in Alzheimer's disease ventricular CSF.
AU Lovell M.A.; Gabbita S.P.; Markesbery W.R.
CS Dr. W.R. Markesbery, 101 Sanders-Brown Building, University of Kentucky, Lexington, KY 40536-0230, United States
SO Journal of Neurochemistry, 1999) 72/2 (771-776).
Refs: 44
ISSN: 0022-3042 CODEN: JONRA
CY United States
DT Journal; Article
FS 008 General Pathology and Pathological Anatomy
009 Neurology and Neurosurgery
LA English
SL English

L5 ANSWER 4 OF 4 LIFESCI COPYRIGHT 2002 CSA
AN 1999:98887 LIFESCI
TI Increased DNA Oxidation and Decreased Levels of Repair Products in Alzheimer's Disease Ventricular CSF
AU Lovell, M.A.; Gabbita, S.P.; Markesbery, W.R.*
CS 101 Sanders-Brown Building, University of Kentucky, Lexington, KY 40536-0230, USA
SO Journal of Neurochemistry [J. Neurochem.], (19990200) vol. 72, no. 2, pp. 771-776.
ISSN: 0022-3042.
DT Journal
FS N3
LA English
SL English

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10 OF 46 CAPLUS COPYRIGHT 2002 ACS
AN 1999:526234 CAPLUS
DN 131:308477
TI Fluorescent **labelling** of closely-spaced aldehydes induced in **DNA** by **bleomycin-Fe**(III)
AU Chakrabarti, S.; Mahmood, A.; Makrigiorgos, G. M.
CS Joint Center for Radiation Therapy and Dana Farber Cancer Institute, Harvard Medical School, Boston, MA, 02215, USA
SO International Journal of Radiation Biology (1999), 75(8), 1055-1065
CODEN: IJRBE7; ISSN: 0955-3002
PB Taylor & Francis Ltd.
DT Journal
LA English
AB The purpose of this study was to test the ability of two novel fluorescent reagents fluorescent aldehyde-reactive probe (FARP) and FARPhc, to **label** aldehyde-contg. sites (principally abasic sites) generated in **DNA** by the radiomimetic drug bleomycin, and to use fluorescent energy transfer from FARPhc (donor) to FARP (acceptor) to quantitate such closely-spaced sites. FARPhc, 7-hydroxycoumarin-3-carboxylic acid (((((amino-oxymethyl) carbonyl) hydrazino) carbonylethyl) amide) was synthesized with a protocol similar to the one recently reported for FARP (a fluorescein-based probe). Both FARPhc and FARP form stable oxime bonds with the open-chain aldehydes generated upon acidic depurination of **DNA**. Plasmid **DNA** exposed to **bleomycin-Fe**(III)-ascorbate undergoes extensive strand breakage, and upon subsequent reaction with FARPhc and/or FARP it becomes fluorescently **labeled**, indicating the generation of aldehyde-contg. sites. The binding of the probes to calf thymus or plasmid **DNA** results in significant fluorescent energy transfer among closely-spaced fluorophores, as revealed by the fluorescence increase following digestion of fluorescently **labeled** samples with nuclease P1. The fluorescence quenching is most evident when both FARPhc and FARP are used simultaneously to trap aldehyde sites. When single-stranded **oligonucleotides** engineered to contain either one or two closely spaced bleomycin binding sites are exposed to bleomycin and then fluorescently **labeled**, the **oligonucleotides** demonstrate significantly increased fluorescent energy transfer with two binding sites indicating a dependence of aldehyde site generation and clustering on the local sequence of a single strand. In conclusion, a new detection method for **DNA** damage induced by bleomycin following fluorescent **labeling** of aldehyde group-contg. sites (FLAGS) and their clustering via fluorescent energy transfer is demonstrated. The method is applicable to any form of **DNA**. This work may lead to a general approach for the quantification of multiply damaged sites in **DNA**, a subset of **DNA** lesions that may have major biol. significance.
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L3 ANSWER 8 OF 16 MEDLINE DUPLICATE 3
AN 97105898 MEDLINE
DN 97105898 PubMed ID: 8948646
TI Chemical methods of DNA and RNA fluorescent labeling.
AU Proudnikov D; Mirzabekov A
CS Engelhardt Institute of Molecular Biology, Moscow, Russia.
SO NUCLEIC ACIDS RESEARCH, (1996 Nov 15) 24 (22) 4535-42.
Journal code: 08L; 0411011. ISSN: 0305-1048.
CY ENGLAND: United Kingdom
DT Journal; Article; (JOURNAL ARTICLE)
LA English
FS Priority Journals
EM 199701
ED Entered STN: 19970219
Last Updated on STN: 19970219
Entered Medline: 19970117
AB Several procedures have been described for fluorescent labeling of DNA and RNA. They are based on the introduction of aldehyde groups by partial depurination of DNA or oxidation of the 3'-terminal ribonucleoside in RNA by sodium periodate. Fluorescent labels with an **attached** hydrazine group are efficiently coupled with the aldehyde groups and the hydrazone bonds are stabilized by reduction with sodium cyanoborohydride. Alternatively, DNA can be quantitatively split at the depurinated sites with ethylenediamine. The aldimine bond between the aldehyde group in depurinated DNA or oxidized RNA and ethylenediamine is stabilized by reduction with sodium cyanoborohydride and the primary amine group introduced at these sites is used for **attachment** of isothiocyanate or succinimide derivatives of fluorescent dyes. The fluorescent DNA labeling can be carried out either in solution or on a reverse phase **column**. These procedures provide simple, inexpensive methods of multiple **DNA labeling** and of introducing one fluorescent dye molecule per RNA, as well as quantitative DNA fragmentation and incorporation of one label per fragment. These methods of fluorophore **attachment** were shown to be efficient for use in the hybridization of labeled RNA, DNA and DNA fragments with oligonucleotide microchips.

L3 ANSWER 10 OF 16 MEDLINE DUPLICATE 4
AN 94057384 MEDLINE
DN 94057384 PubMed ID: 8238885
TI Biotinylation of **DNA** on membrane **supports**: a procedure for preparation and easy control of **labeling** of nonradioactive single-stranded **nucleic** acid probes.
AU Didenko V V
CS Department of Immunology, Institute of Transplantology and Artificial Organs, Moscow, Russia.
SO ANALYTICAL BIOCHEMISTRY, (1993 Aug 15) 213 (1) 75-8.
Journal code: 4NK; 0370535. ISSN: 0003-2697.
CY United States
DT Journal; Article; (JOURNAL ARTICLE)
LA English
FS Priority Journals
EM 199312
ED Entered STN: 19940117
Last Updated on STN: 19940117
Entered Medline: 19931203
AB We have used M13 single-stranded DNA **bound** by uv to small pieces of nylon membrane for the synthesis of biotinylated single-stranded DNA probes. The labeling method requires a large fragment of DNA polymerase I and random hexanucleotides. There is no need for previous linearization of the template. The clean probe is removed from the membrane by a single wash step. The synthesized probe is completely free of unincorporated precursors. This makes possible the easy control of the reaction of incorporation of biotinylated analogues into the probe by simple staining on the filter, thus allowing evaluation of the efficiency of labeling. The DNA membrane can be stored for reuse. With the procedure described it is possible to biotinylate many DNA fragments in parallel, simultaneously controlling the efficiency of labeling in a time- and cost-saving manner.